# Morphological variability of black bullhead Ameiurus melas in four non-native European populations 

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#### Abstract

External morphology in black bullhead Ameiurus melas, a fish species considered to have high invasive potential, was studied in its four non-native European populations (British, French, Italian and Slovak). The aim of this study was to examine this species' variability in external morphology, including ontogenetic context, and to evaluate its invasive potential. Specimens from all non-native populations reached smaller body size compared to individuals from native populations. Juvenile A. melas were found to have a relatively uniform body shape regardless of the population's origin, whereas adults developed different phenotypes depending upon location. Specimens from the U.K., Slovak and French populations appeared to be rather similar to each other, whereas the Italian population showed the most distant phenotype. This probably results from the different thermal regime in the Italian habitat. Ameiurus melas from non-native European populations examined in this study showed some potential to alter the body shape both within and between populations. The phenotypic plasticity of A. melas, however, was not found to be as significant as in other invasive fish species. The results suggest that morphological variability itself is not necessarily essential for invasive success. The invasiveness of $A$. melas is therefore probably favoured by variations in its life-history traits and reproduction variables, together with some behavioural traits (e.g. voracious feeding and parental care) rather than by phenotypic plasticity expressed in external morphology.

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Key words: external morphology; geometric morphometrics; growth changes; invasive species; ontogeny.

## INTRODUCTION

Variability in growth, development and maturation creates a variety of body shapes within a species (Cadrin, 2000) that, along with ecological interactions of organisms, are directly or indirectly influenced by environmental conditions (Norton et al., 1995). Some ontogenetic features are obscured or displaced by subsequent development, but

[^0]others persist as a record of an individual's life history. Moreover, interactive effects of environment, selection and genetics on individual ontogenies produce variation in the morphology of a species (Cadrin, 2000). Particularly in the case of introduced species, the ability to adapt to a novel environment plays an important role in determining their invasion success (Sakai et al., 2001; McMahon, 2002). Phenotypic plasticity allows the same genotype to produce a variety of phenotypes in response to different local conditions (Komers, 1997; Schlichting \& Pigliucci, 1998). In other words, phenotypic plasticity allows introduced species to respond effectively to environmental changes by modification of morphology, reproduction or survival that mitigate the effects of environmental variation (Stearns, 1983; Meyer, 1987). The detailed examination of external morphology of rapidly dispersing species usually shows a high degree of phenotypic plasticity (Balážová-Ľavrinčíková \& Kováč, 2007; Tomeček et al., 2007; Záhorská et al., 2009), and morphometry is one of the most easily accessible means of assessing a species' environmental plasticity (Kováč et al., 1999).

The black bullhead Ameiurus melas (Rafinesque 1820) was introduced to Europe in the late 19th and early 20th centuries from North America and is now established in many European countries (Wheeler, 1978; Copp et al., 2005). Ameiurus melas possesses several characteristics associated with successful invaders (Ribeiro et al., 2008), such as high reproductive potential, parental care, omnivory, aggressive behaviour and considerable tolerance to water pollution, turbidity, low oxygen concentration, elevated temperatures and a range of pH values (Scott \& Crossman, 1973; Karp \& Tyus, 1990; Brown et al., 1999; Vila-Gispert et al., 2005; Leunda et al., 2008; Ribeiro et al., 2008; Novomeská \& Kováč, 2009). The species, however, has received little attention regarding its external morphology and phenotypic plasticity, attributes that may be important for successful invasion (Záhorská et al., 2009). As geographic variation in morphometry has been used to discriminate local forms of fishes (Cadrin, 2000), the aim of this study was to examine variability in the external morphology of A. melas from non-native populations in four European countries (France, Italy, Slovakia and U.K.). Evaluated within an ontogenetic context, morphological variability is discussed in terms of this species' invasive potential.

## MATERIALS AND METHODS

Specimens of A. melas were collected from four European sites (Fig. 1): Tom's Pond $\left(51^{\circ} 42^{\prime} 26^{\prime \prime} \mathrm{N} ; 0^{\circ} 10^{\prime} 58^{\prime \prime} \mathrm{E}\right)$, Essex, south-east U.K. $(n=163)$; Brière Marsh $\left(47^{\circ} 22^{\prime} \mathrm{N}\right.$; $2^{\circ} 11^{\prime}$ W), River Loire drainage, north-west France ( $n=93$ ); Lake Trasimeno ( $43^{\circ} 05^{\prime} 21^{\prime \prime} \mathrm{N}$; $\left.12^{\circ} 09^{\prime \prime} 18^{\prime \prime} \mathrm{E}\right)$, Umbria region, central Italy $(n=342)$ and from an oxbow of the River Váh $\left(47^{\circ} 48^{\prime} 30^{\prime \prime} \mathrm{N} ; 18^{\circ} 07^{\prime} 30^{\prime \prime} \mathrm{E}\right.$ ) in Komárno, south-western Slovakia $n=231$; evaluation of the last is based on reanalysed data already published by Novomeská et al. (2010)]. The fish were killed with an overdose of anaesthetic and immediately chilled to freezing for subsequent analysis in the laboratory. Once defrosted, all specimens were photographed from the lateral and dorsal perspectives using a Nikon D40 digital camera (www.nikon.com), with 32 morphometric characters (Fig. 2), including standard length ( $L_{\mathrm{S}}$ ) and total length $\left(L_{T}\right)$, subsequently measured using Impor 2.31 E software (Kvant s.r.o.; www.kvant.sk). To examine the patterns of relative growth (ontogenetic aspect), raw values from 30 morphometric characters were plotted against $L_{\mathrm{S}}$ (Kováč et al., 1999). Inter-population comparisons of the distance-based measurements (Table I) were based on values of these 30 morphometric characters expressed in $\% L_{\mathrm{S}}$ (Snedecor, 1946). All specimens (females,


Fig. 1. Sampling sites of four non-native European Ameiurus melas populations (U.K., French, Italian and Slovak).
males and juveniles) were analysed together as no sexual dimorphism has previously been found in A. melas (Novomeská et al., 2010).

Using triple-regression analysis (TRA), the development of body proportions was assumed not to deviate from isometry (linear regression; the null hypothesis $H_{0}$ ), with gradual allometry (quadratic regression; $H_{\mathrm{a}_{1}}$ ) and isometry with an abrupt change (split linear regression; $H_{\mathrm{a}_{2}}$ ) being the alternative hypotheses. In isometric development, the morphometric characters develop in proportion to $L_{\mathrm{S}}$, while in allometric development the growth rate is always different from that of $L_{\mathrm{S}}$. In isometric with abrupt change, a character develops isometrically up to a specific $L_{\mathrm{S}}$, then changes to develop allometrically within a short $L_{\mathrm{S}}$ interval and subsequently shifts to another isometric relationship. Shifts, which separated two intervals of isometric growth, were indicated by a breakpoint between two linear regressions (Nickerson et al., 1989). Finally, the model that provided the best fit was assessed using the $F$-test (Sokal \& Rohlf, 1981; see Tables II and III).

The results of TRA (predominant type of development in each population) were used in correspondence analysis (CA) to find out multivariate mutual relationship between populations. The findings were visualized as CA scores and loadings.

As morphological variability can be explained by transformations of homologous features in co-ordinate space (Thompson, 1917), geometrical shape analysis of landmarks [co-ordinates-based measurements in a two-dimensional space (Katina, 2003)] was used. A wide range of body shapes can be described as regular compressions or elongations in a rectangular co-ordinate system (Thompson, 1917), and thus coordinates of 18 landmarks (Fig. 2) in each specimen were marked in the ImporPro 3.2 software (Kvant s.r.o., Bratislava, Slovakia) and evaluated in R software ( R Development Core Team; www.r-project.org) with programme routines as established by Katina (2007). Subsequently, the landmark-point co-ordinates were transformed to Procrustes shape co-ordinates using generalized Procrustes analysis (Dryden \& Mardia, 1999). First, the centroid of each form was found, and its root-mean-square distance to the landmarks (i.e. centroid size, $C_{\mathrm{S}}$ ) was identified. Second, the forms were rescaled to $C_{\mathrm{S}}=1$, the centroids superimposed and the forms rotated around one another about the common centroid until the sum-of-squared distances between corresponding landmarks achieved a minimum over all such rotations. This resulted in Procrustes shape co-ordinates.

| Character | U.K. |  |  |  | FRA |  |  |  | ITA |  |  |  | SVK |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Mean | Min | Max | c.v. | Mean | Min | Max | c.v. | Mean | Min | Max | c.v. | Mean | Min | Max | c. |
| Total length (mm) | 113.7 | 72.7 | 228.0 | 25.0 | 152.5 | 57.7 | 231.0 | $30 \cdot 0$ | 182.1 | 34.5 | 261.3 | 24.1 | $120 \cdot 2$ | 20.8 | 219.7 | 33.9 |
| $L_{\mathrm{S}}(\mathrm{mm})$ | 94.5 | $60 \cdot 3$ | 193.8 | $26 \cdot 0$ | $130 \cdot 5$ | 49.1 | $200 \cdot 8$ | $30 \cdot 6$ | 158.3 | 29.5 | 229.4 | 24.3 | 101.8 | 17.6 | 184.9 | 34.0 |
| $\%$ of $L_{S}$ |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 1. Head length | 29.0 | 23.5 | $35 \cdot 8$ | 8.5 | 25.6 | 23.4 | $33 \cdot 2$ | 7.6 | 29.8 | $25 \cdot 8$ | 35.7 | $5 \cdot 5$ | 27.2 | $20 \cdot 1$ | 34.4 | 8.2 |
| 2. Preorbital distance | 7.2 | 4.8 | $10 \cdot 3$ | 13.9 | $7 \cdot 4$ | $5 \cdot 5$ | 9.9 | $10 \cdot 9$ | 9.3 | 6.0 | $12 \cdot 6$ | 11.7 | $7 \cdot 1$ | 4.8 | 9.0 | $12 \cdot 6$ |
| 3. Eye diameter | 3.8 | 2.4 | $5 \cdot 6$ | 14.9 | 3.0 | $2 \cdot 3$ | 4.3 | $15 \cdot 1$ | $2 \cdot 5$ | 1.7 | 5. 1 | 18.1 | 4.0 | 2.8 | 6.9 | 18.7 |
| 4. Postorbital distance | 18.3 | 12.2 | 22.4 | 11.6 | 17.3 | 13.8 | 20.8 | 9.4 | 18.1 | 14.7 | 22.3 | 7.7 | $16 \cdot 8$ | 13.7 | 20.9 | 7.2 |
| 5. Head depth | 22.0 | $15 \cdot 8$ | 25.9 | 7.5 | 21.9 | 17.5 | 26.7 | 8.8 | 20.7 | 16.5 | 26.4 | $10 \cdot 1$ | 21.5 | 18.3 | 25.9 | $7 \cdot 0$ |
| 6. Predorsal distance | $41 \cdot 6$ | 33.0 | 48.6 | 4.4 | 41.8 | 38.6 | 45.9 | 4.0 | $40 \cdot 6$ | 36.4 | 45.5 | 3.7 | 41.0 | 35.5 | 45.9 | 4.5 |
| 7. Prepelvic distance | $50 \cdot 1$ | 33.2 | 55.3 | 5. 1 | $50 \cdot 2$ | 43.9 | 55.0 | 4.2 | 51.0 | 46.4 | 58.2 | $3 \cdot 8$ | 49.9 | $45 \cdot 0$ | 57.4 | 3.9 |
| 8. Preanal distance | 63.5 | 52.4 | $75 \cdot 8$ | $5 \cdot 2$ | 64.1 | $55 \cdot 5$ | 74.1 | 4.0 | 63.7 | $56 \cdot 7$ | 71.5 | $3 \cdot 0$ | $62 \cdot 1$ | 58.8 | 71.2 | $2 \cdot 8$ |
| 9. Postdorsal distance | $56 \cdot 2$ | 44.6 | $62 \cdot 9$ | 4.9 | 56.5 | $52 \cdot 1$ | 63.6 | 4.0 | $55 \cdot 5$ | 50.0 | $60 \cdot 3$ | $3 \cdot 1$ | 56.6 | 50.8 | 62.7 | $3 \cdot 3$ |
| 10. V-A distance | 13.8 | 8.7 | $22 \cdot 3$ | 17.0 | 14.4 | $10 \cdot 0$ | $20 \cdot 6$ | 13.9 | 13.3 | 7.8 | 18.7 | 11.6 | 12.7 | 9.6 | 15.5 | 9.5 |
| 11. D-A distance | 37.4 | $30 \cdot 5$ | 42.7 | $5 \cdot 5$ | 38.4 | 33.4 | 44.1 | $5 \cdot 5$ | $36 \cdot 0$ | 32.2 | 42.9 | 4.6 | 36.5 | 32.0 | 40.4 | 3.8 |
| 12. D-adip distance | 35.7 | 26.2 | 44.4 | 8.0 | 37.2 | $30 \cdot 2$ | $46 \cdot 3$ | 7.4 | 37.3 | 32.0 | 44.5 | 5.7 | 36.3 | $30 \cdot 3$ | $43 \cdot 1$ | 5.4 |
| 13. Adip-A distance | 26.3 | 20.0 | $30 \cdot 6$ | 7.2 | 27.7 | 22.9 | $31 \cdot 1$ | $6 \cdot 2$ | 25.3 | 20.3 | 32.2 | 6.9 | 26.9 | $22 \cdot 6$ | $30 \cdot 7$ | $5 \cdot 6$ |
| 14. Adip-posterior of A distance | 22.9 | 16.8 | 27.9 | 7.9 | 22.5 | 18.1 | 27.9 | 8.5 | 20.8 | 13.2 | 26.7 | 7.4 | 23.0 | 18.6 | 27.3 | 6. 8 |
| 15. Posterior of adip-C base | 19.2 | 14.9 | 23.0 | 7.5 | 18.7 | 15.4 | 21.6 | 6.5 | 17.6 | 14.2 | 23.5 | 7.1 | 19.8 | 16.4 | $23 \cdot 6$ | 5.7 |
| 16. Caudal-peduncle length | 17.0 | 13.8 | 19.6 | 6.5 | 16.4 | 14.2 | 19.7 | 7.4 | 16.1 | 11.8 | 20.6 | 7.7 | 16.4 | 13.7 | 19.5 | $6 \cdot 1$ |
| 17. Caudal-peduncle depth | 14.7 | 11.3 | 17.7 | 6.6 | 14.5 | $12 \cdot 7$ | 16.6 | 6.3 | 13.8 | $10 \cdot 6$ | 17.8 | 8.6 | 14.6 | 11.8 | 17.2 | 5.9 |
| 18. Minimum body depth | 14.2 | 10.7 | $16 \cdot 2$ | $5 \cdot 6$ | 13.7 | 12.3 | 15.8 | 5.4 | 12.5 | 10. 1 | $16 \cdot 2$ | 7.1 | 14.1 | 11.2 | 16.4 | 5.9 |
| 19. Body depth | 29.7 | 18.9 | $40 \cdot 6$ | $10 \cdot 4$ | 29.9 | 24.4 | 39.1 | 11.3 | 26.5 | 18.7 | 37.3 | 12.6 | 29.8 | 24.3 | $36 \cdot 2$ | 8.6 |
| 20. D height | 19.9 | 13.2 | 25.8 | 9.2 | 17.3 | $12 \cdot 1$ | 21.0 | 11.1 | 16.0 | 10.7 | 26.4 | $13 \cdot 1$ | 19.3 | 11.7 | 23.3 | 7.8 |
| 21. V height | 13.4 | 8.4 | 17.3 | 14.2 | 13.0 | 8.4 | 17.5 | 13.6 | 13.2 | 9.0 | 16.7 | 9.2 | 13.8 | 9.4 | 18.3 | 9.6 |
| 22. A height | 11.5 | 7.7 | $15 \cdot 8$ | 14.2 | 11.2 | 6.0 | $15 \cdot 8$ | 16.9 | 11.2 | 7.1 | 14.9 | 13.3 | 13.3 | 9. 1 | 17.4 | 11.2 |
| 23. C height | 28.5 | 20.6 | 38.3 | 12.7 | 24.8 | 18.8 | 31.1 | $10 \cdot 3$ | 21.3 | $15 \cdot 0$ | $30 \cdot 0$ | 14.1 | 25.2 | 16.3 | 31.5 | 11.2 |
| 24. D-base length | 9.3 | 6.8 | 13.7 | 11.0 | 9.0 | 6.3 | 11.2 | $10 \cdot 3$ | 9. 2 | 5.8 | 13.5 | 11.2 | 9.3 | 7.0 | 15.6 | 13.2 |
| 25. Adip-base length | $11 \cdot 0$ | 6.8 | 15.4 | $16 \cdot 1$ | $10 \cdot 3$ | 7.0 | $15 \cdot 0$ | 14.9 | $10 \cdot 8$ | 6.7 | $15 \cdot 0$ | 13.9 | 11.7 | 8.1 | 22.6 | 13.7 |
| 26. A-base length | 22.2 | 14.3 | 26.7 | $10 \cdot 6$ | 23.0 | 18.5 | 27.3 | 7.8 | 22.9 | 17.4 | 26.6 | 6.0 | 23.7 | $16 \cdot 2$ | 27.6 | 6.0 |
| 27. C length | 20.9 | 15.9 | 23.3 | 6.8 | 17.3 | $15 \cdot 0$ | 21.7 | 8.4 | $15 \cdot 2$ | 11.3 | 20.9 | 9.2 | 18.4 | 14.0 | 21.3 | 6.5 |
| 28. P length | 15.5 | 12. 1 | 18.6 | 7.3 | 14.1 | 10.7 | 18.1 | 9.9 | 13.0 | 9.0 | 19.5 | 12.4 | $15 \cdot 7$ | $10 \cdot 8$ | 21.6 | 8.9 |
| 29. Interorbital distance | 18.2 | 13.6 | 21.7 | 6.9 | 19.2 | $16 \cdot 6$ | 24.0 | 7.9 | 18.4 | 14.7 | $23 \cdot 6$ | 9.3 | 19.5 | 15.7 | 23.3 | 6.7 |
| 30. Head width | $26 \cdot 2$ | 21.5 | 29.3 | 4.1 | 26.1 | 23.0 | 29.9 | 4.9 | 24.8 | $20 \cdot 2$ | $30 \cdot 3$ | 8.3 | 27.2 | 24.2 | 33.4 | $5 \cdot 0$ |

Table II. Triple-regression analysis for morphometric characters (see Table I) of Ameiurus melas from U.K. and France

| Character | U.K. |  |  |  |  |  |  |  |  | France |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $F(\mathrm{Q} / \mathrm{L})$ | $P$ | $F(\mathrm{~S} / \mathrm{Q})$ | $P$ | $F(\mathrm{~S} / \mathrm{L})$ | $P$ | Fit | BP | s.e. | $F(\mathrm{Q} / \mathrm{L})$ | $P$ | $F$ (S/Q) | $P$ | $F(\mathrm{~S} / \mathrm{L})$ | $P$ | Fit | BP | S.E |
| 1. Head length | 2.77 | NS | -2. 23 | NS | 0. 24 | NS | L |  |  | 4.08 | <0.05 | $-0.87$ | NS | 1.56 | NS | Q |  |  |
| 2. Preorbital distance | 2. 59 | NS | -6.17 | NS | $-1.84$ | NS | L |  |  | 0. 56 | NS | 0. 38 | NS | 0.47 | NS | L |  |  |
| 3. Eye diameter | 1. 22 | NS | $-12.86$ | NS | $-5.87$ | NS | L |  |  | 3.75 | NS | -15.28 | NS | -6.11 | NS | L |  |  |
| 4. Postorbital distance | $0 \cdot 68$ | NS | $1 \cdot 17$ | NS | 0.93 | NS | L |  |  | 2. 14 | NS | -1.38 | NS | 0.35 | NS | L |  |  |
| 5. Head depth | 0. 69 | NS | -0. 20 | NS | 0. 24 | NS | L |  |  | 0.98 | NS | 2. 14 | NS | 1.56 | NS | L |  |  |
| 6. Predorsal distance | 8.33 | $<0.01$ | $-29.16$ | NS | $-11.20$ | NS | Q |  |  | 9.48 | <0.01 | 6.37 | $<0.05$ | 8.21 | <0.01 | S | 157.97 | $6 \cdot 29$ |
| 7. Prepelvic distance | 4. 18 | <0.05 | -2.44 | NS | 0. 82 | NS | Q |  |  | $0 \cdot 00$ | NS | $0 \cdot 04$ | NS | $0 \cdot 02$ | NS | L |  |  |
| 8. Preanal distance | 1.81 | NS | 3. 11 | NS | 2.47 | NS | L |  |  | $0 \cdot 62$ | NS | 1.00 | NS | 0.81 | NS | L |  |  |
| 9. Postdorsal distance | 2.71 | NS | 5.38 | $<0.05$ | 4.08 | $<0.05$ | S | 190.70 | 1.63 | $0 \cdot 00$ | NS | 1.39 | NS | $0 \cdot 69$ | NS | L |  |  |
| 10. V-A distance | 8.82 | <0.01 | -31.54 |  | $-12.26$ | NS | Q |  |  | $2 \cdot 68$ | NS | -28.31 | NS | -13.25 | NS | L |  |  |
| 11. D-A distance | $0 \cdot 69$ | NS | 10.46 | $<0.01$ | 5. 59 | <0.05 | S | 182.85 | 3.45 | 0.30 | NS | 6.74 | $<0.05$ | 3.53 | NS | L |  |  |
| 12. D -adip distance | 0. 00 | NS | 0.75 | NS | 0. 37 | NS | L |  |  | 0. 15 | NS | 1.47 | NS | 0.81 | NS | L |  |  |
| 13. Adip-A distance | 3. 70 | NS | $-1.80$ | NS | 0.91 | NS | L |  |  | $0 \cdot 22$ | NS | 1.34 | NS | 0.78 | NS | L |  |  |
| 14. Adip-posterior of A dist. | 0. 47 | NS | 7.01 | $<0.01$ | 3.75 | NS | L |  |  | 0. 12 | NS | $3 \cdot 07$ | NS | 1.60 | NS | L |  |  |
| 15. Posterior of adip-C base | 0.77 | NS | 13.93 | $<0.01$ | 7.38 | $<0.01$ | S | 185.81 | 3.07 | 3.58 | NS | 1.87 | NS | 2.75 | NS | L |  |  |
| 16. Caudal-peduncle length | 0. 43 | NS | 7.81 | $<0.01$ | 4. 13 | $<0.05$ | S | 177.21 | 6.55 | 1.02 | NS | 2. 19 | NS | 1.61 | NS | L |  |  |
| 17. Caudal-peduncle depth | 0. 99 | NS | 18.36 | <0.01 | 9. 73 | <0.01 | S | 185.79 | 2.69 | 0. 20 | NS | 6. 22 | <0.05 | 3.22 | NS | L |  |  |
| 18. Minimum body depth | 0. 00 | NS | 0.30 | NS | 0. 15 | NS | L |  |  | 0.54 | NS | 5.42 | <0.05 | 3.00 | NS | L |  |  |
| 19. Body depth | 6.75 | <0.05 | -23.58 | NS | -8.93 | NS | Q |  |  | 0. 10 | NS | 1.71 | NS | 0.91 | NS | L |  |  |
| 20. D height | 4. 81 | $<0.05$ | 2.77 | NS | $3 \cdot 82$ | NS | Q |  |  | $5 \cdot 82$ | <0.05 | -29.92 | NS | $-13.05$ | NS | Q |  |  |
| 21. V height | $2 \cdot 82$ | NS | 1.01 | NS | 1.92 | NS | L |  |  | $0 \cdot 06$ | NS | 0.36 | NS | 0.21 | NS | L |  |  |
| 22. A height | $2 \cdot 08$ | NS | -9.33 | NS | -3.69 | NS | L |  |  | 0. 19 | NS | $-18.62$ | NS | -9.23 | NS | L |  |  |
| 23. C height | 3.71 | NS | 0.61 | NS | 2. 15 | NS | L |  |  | 10.21 | <0.01 | 1.74 | NS | $6 \cdot 02$ | <0.05 | Q |  |  |
| 24. D-base length | $6 \cdot 12$ | <0.05 | -12.82 | NS | -3.61 | NS | Q |  |  | 8.51 | <0.01 | -29. 02 | NS | -11.68 | NS | Q |  |  |
| 25. Adip-base length | 0. 04 | NS | -21.59 | NS | $-10.77$ | NS | L |  |  | 0. 28 | NS | $-18.27$ | NS | -9.02 | NS | L |  |  |
| 26. A-base length | 1. 11 | NS | 6. 19 | $<0.05$ | 3.67 | NS | L |  |  | $0 \cdot 65$ | NS | -32.56 | NS | -16.07 | NS | L |  |  |
| 27. C length | 28.56 | <0.01 | -3.37 | NS | 12. 19 | <0.01 | Q |  |  | 14.93 | <0.01 | -2.76 | NS | 5.77 | <0.05 | Q |  |  |
| 28. P length | 2.41 | NS | 4.28 | $<0.05$ | 3. 37 | NS | L |  |  | 1.96 | NS | 1.62 | NS | 1.80 | NS | L |  |  |
| 29. Interorbital distance | 0. 29 | NS | 11.41 | $<0.01$ | $5 \cdot 86$ | <0.05 | S | $186 \cdot 15$ | 3.01 | 9. 20 | $<0.01$ | 0.96 | NS | 5.08 | $<0.05$ | Q |  |  |
| 30. Head width | 0 | NS | 1.85 | NS | 0.92 | NS | L |  |  | 6.35 | $<0.05$ | $2 \cdot 02$ | NS | $4 \cdot 22$ | <0.05 | Q |  |  |



Fig. 2. Schematic illustration of the distance-based morphometric characters (__) and the 18 landmarks ( used in geometrical shape analysis of Ameiurus melas. (a) Lateral and (b) dorsal view: total length (1-2), standard length $(1-3)$, head length $(1-4)$, preorbital distance $(1-5)$, eye diameter (5-6), postorbital distance (6-4), head depth $(7-8)$, predorsal distance $(1-9)$, prepelvic distance $(1-10)$, pre-anal distance ( $1-11$ ), post-dorsal distance $(12-3), \mathrm{V}-\mathrm{A}$ distance $(10-11)$, $\mathrm{D}-\mathrm{A}$ distance $(9-11)$, $\mathrm{D}-$ adip distance ( $9-13$ ), adip-A distance ( $13-11$ ), adip-posterior of A distance (13-14), posterior of adip-C base ( $15-3$ ), caudal-peduncle length ( $14-3$ ), caudal-peduncle depth ( $14-16$ ), minimum body depth ( $17-18$ ), body depth $(9-19)$, D height $(9-20)$, V height ( $10-21$ ), A height ( $22-23$ ), C height ( $24-25$ ), D-base length (9-12), adip-base length (13-15), A-base length (11-14), C length (2-3), P length (26-27), interorbital distance (28-29), head width (30-31) [dorsal (D), pectoral (P), pelvic (V), anal (A) and caudal (C) fins and adipose fin (adip)].

To explore the nature of size and shape (form) variability, form space was decomposed into: (1) size-related and (2) size-adjusted sub-spaces. In the size-related sub-space, the allometric pattern (shape changes during growth) was analysed via form-space principal component analysis (PCA; relative warp analysis, RWA). In the form-space PCA, the approximate tangent co-ordinates (centred Procrustes shape co-ordinates) were augmented by $\ln C_{\mathrm{S}}$, and variability was decomposed into orthogonal components of size-and-shape variation, which were examined. The first principal component ( PC 1 ) usually reflects changes in external morphology associated with growth changes (allometry), which is measured by Pearson product moment correlation coefficient of PC1 scores and $\log _{10} C_{\mathrm{S}}$. In the size-adjusted sub-space, the nonallometric pattern was analysed via size-adjusted PCA. In this form-space decomposition, size-adjusted variability is orthogonal (perpendicular) to the growth direction, geometrically speaking, and both sub-spaces are independent; thus, the allometric and non-allometric biological signals are decomposed and analysed separately. In both PCA methods, the first two PCs accounted for a sufficient amount of variability ( $97.53 \%$ ).

To explore the nature of differences between the populations, a linear discriminant analysis (LDA) was used. As in PCA, the variability was decomposed into two components: (1) size-related, i.e. analysed with shape-space LDA and (2) size-adjusted, i.e. analysed with size-adjusted shape-space LDA. In both LDA methods, the first two linear discriminants
Table III. Triple-regression analysis for morphometric characters of Ameiurus melas from Italy and Slovakia (see Table II)

| Character | Italy |  |  |  |  |  |  |  |  | Slovakia |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $F(\mathrm{Q} / \mathrm{L})$ | $P$ | $F$ (S/Q) | $P$ | $F$ (S/L) | $P$ | Fit | BP | s.e. | $F(\mathrm{Q} / \mathrm{L})$ | $P$ | $F$ (S/Q) | $P$ | $F$ (S/L) | $P$ | Fit | BP | S.E |
| 1. Head length | 6.22 | <0.05 | -4.85 | NS | 0. 62 | NS | Q |  |  | 0.34 | NS | 11.02 | <0.01 | $5 \cdot 68$ | $<0.05$ | S | 151 -12 | 4.4 |
| 2. Preorbital distance | $0 \cdot 62$ | NS | $-17.67$ | NS | $-8.54$ | NS | L |  |  | 4.30 | <0.05 | $-21.00$ | NS | $-8.55$ | NS | Q |  |  |
| 3. Eye diameter | 0. 85 | NS | $-15.62$ | NS | $-7.40$ | NS | L |  |  | 11.48 | <0.01 | $1 \cdot 18$ | NS | 6.34 | <0.05 | Q |  |  |
| 4. Postorbital distance | 10.10 | $<0.01$ | $7 \cdot 16$ | <0.01 | 8.72 | $<0.01$ | S | 172.27 | 7.65 | $0 \cdot 00$ | NS | 0. 10 | NS | 0.05 | NS | L |  |  |
| 5. Head depth | 22.97 | $<0.01$ | -8. 52 | NS | $6 \cdot 90$ | <0.01 | Q |  |  | 5.76 | <0.05 | -2. 26 | NS | 1.71 | NS | Q |  |  |
| 6. Predorsal distance | 15.79 | $<0.01$ | 1. 13 | NS | 8.46 | $<0.01$ | Q |  |  | 1.68 | NS | 12.35 | <0.01 | 7.05 | $<0.01$ | S | 150.09 | 4.0 |
| 7. Prepelvic distance | 23.51 | $<0.01$ | $-10.24$ | NS | 6. 24 | $<0.05$ | Q |  |  | 6.96 | <0.01 | 0. 72 | NS | 3.83 | NS | Q |  |  |
| 8. Preanal distance | 32.70 | <0.01 | $-18.97$ | NS | $5 \cdot 90$ | <0.05 | Q |  |  | 11.94 | <0.01 | $7 \cdot 07$ | <0.01 | 9.67 | <0.01 | S | 88.52 | 7.8 |
| 9. Postdorsal distance | $10 \cdot 62$ | $<0.01$ | 2.02 | NS | 6.34 | <0.05 | Q |  |  | $5 \cdot 82$ | <0.05 | -2.21 | NS | 1.76 | NS | Q |  |  |
| 10. V-A distance | 2. 20 | NS | -0.65 | NS | 0.78 | NS | L |  |  | 3.68 | NS | -11.39 | NS | -3.95 | NS | L |  |  |
| 11. D-A distance | 9.38 | <0.01 | $-1.49$ | NS | 3. 91 | <0.05 | Q |  |  | 5.63 | <0.05 | $5 \cdot 37$ | $<0.05$ | 5.56 | $<0.05$ | S | 132.74 | 7.9 |
| 12. D -adip distance | 0. 60 | NS | -0. 14 | NS | 0. 23 | NS | L |  |  | 11.00 | <0.01 | 4.64 | $<0.05$ | 7.91 | $<0.01$ | S | 155.35 | 3.62 |
| 13. Adip-A distance | 4.43 | $<0.05$ | 5. 29 | <0.05 | 4. 89 | <0.05 | S | $166 \cdot 89$ | 10.56 | 16.09 | <0.01 | 7.31 | $<0.01$ | 11.92 | $<0.01$ | S | 86.83 | 6.9 |
| 14. Adip-posterior of A dist. | 9.64 | $<0.01$ | -0. 24 | NS | $4 \cdot 68$ | <0.05 | Q |  |  | 3.41 | NS | 2.76 | NS | 3. 10 | NS | L |  |  |
| 15. Posterior of adip-C base | 3.27 | NS | -2.83 | NS | 0. 20 | NS | L |  |  | 18.29 | <0.01 | 2.97 | NS | 10.71 | $<0.01$ | Q |  |  |
| 16. Caudal peduncle length | 5.83 | <0.05 | 2.99 | NS | 4. 43 | <0.05 | Q |  |  | $0 \cdot 00$ | NS | 8.48 | $<0.01$ | 4.24 | $<0.05$ | S | 153.53 | 4.5 |
| 17. Caudal peduncle depth | 2.65 | NS | -0.23 | NS | 1. 20 | NS | L |  |  | 23.71 | <0.01 | 8.04 | <0.01 | 16.24 | $<0.01$ | S | 106.85 | 5.9 |
| 18. Minimum body depth | $3 \cdot 86$ | NS | 1.98 | NS | 2. 93 | NS | L |  |  | 34.41 | <0.01 | 10.99 | <0.01 | 23.46 | $<0.01$ | S | 98.27 | 4.9 |
| 19. Body depth | 2.34 | NS | 4.66 | <0.05 | 3.51 | NS | L |  |  | $0 \cdot 00$ | NS | 7.47 | $<0.01$ | 3.73 | NS | L |  |  |
| 20. D height | 1.74 | NS | $-329.94$ | NS - | 164.95 | NS | L |  |  | 2. 83 | NS | -0. 56 | NS | $1 \cdot 12$ | NS | L |  |  |
| 21. V-height | 0.89 | NS | -0. 57 | NS | 0. 15 | NS | L |  |  | 0.35 | NS | 1.61 | NS | 0.98 | NS | L |  |  |
| 22. A height | 0. 15 | NS | -25.78 | NS - | $-12 \cdot 82$ | NS | L |  |  | 14.06 | <0.01 | 11.35 | <0.01 | 13.03 | <0.01 | S | 120.47 | 5.6 |
| 23. C height | 0. 25 | NS | 9. 14 | <0.01 | 4. 70 | <0.05 | S | 226.40 | $1 \cdot 13$ | 32.59 | <0.01 | 2.79 | NS | 17.81 | <0.01 | Q |  |  |
| 24. D-base length | 12.64 | <0.01 | 0.51 | NS | 6.56 | <0.05 | Q |  |  | 13.85 | <0.01 | $5 \cdot 32$ | $<0.05$ | 9.72 | $<0.01$ | S | 77.76 | $6 \cdot 3$ |
| 25. Adip-base length | 8.79 | $<0.01$ | $-27.35$ | NS | $-9.64$ | NS | Q |  |  | 0.79 | NS | -2.14 | NS | $-0.67$ | NS | L |  |  |
| 26. A-base length | 13.35 | $<0.01$ | 1.48 | NS | 7.42 | <0.01 | Q |  |  | 0.74 | NS | 3.31 | NS | 2.03 | NS | L |  |  |
| 27. C length | 5.47 | <0.05 | -17.34 | NS | -6.08 | NS | Q |  |  | 2.65 | NS | 1.85 | NS | 2.26 | NS | L |  |  |
| 28. P length | 2. 17 | NS | 0.58 | NS | 1.37 | NS | L |  |  | 16.96 | <0.01 | 6.33 | <0.05 | 11.84 | <0.01 | S | 98.20 | 7.0 |
| 29. Interorbital distance | 20.33 | <0.01 | $-14.60$ | NS | 2. 39 | NS | Q |  |  | $2 \cdot 81$ | NS | 4.67 | <0.05 | 3.76 | NS | L |  |  |
| 30. Head width | 14.41 | $<0.01$ | $-7.55$ | NS | 3. 24 | NS | Q |  |  | 1.50 | NS | 10. 11 | $<0.01$ | 5.83 | $<0.05$ | S | 159.95 | 4.8 |

[^1](LD) covered sufficient amount of variability (90.44 \%). Interpretation of the PCs and LDs followed Mitteroecker et al. (2004) and Katina (2007), where the effects of each PC and LD were visualized (i.e. back-projected into the configuration space) via thin-plate spline (TPS) deformation grids, for both directions of negative (left) and positive (right) change, corresponding to the particular PC and LD direction. To ease the visualization, the differences were magnified two (PCA) or three (LDA) times.

The coefficient of difference ( $C_{\text {dif }}$ ) was used to compare $L_{S}$ as well as the other morphometric characters among the populations examined: $C_{\text {dif }}=\left(x_{2}-x_{1}\right)\left(s_{1}+s_{2}\right)^{-1}$, where $x_{1}$ and $x_{2}$ are the arithmetic averages, and $s_{1}$ and $s_{2}$ are the s.D. Real differences between $90 \%$ of the populations or morphometric characters exist when $C_{\text {dif }}>1.28$ (Mayr et al., 1953). For ontogenetic evaluations, as well as for the inter-population comparisons of distancebased morphometric characters, two sub-samples of 30 specimens with similar range of $L_{\mathrm{S}}$ (i.e. $C_{\text {dif }}<1.28$ ) were selected from each population. These two sub-samples consisted of smallest and largest, or juvenile and adult specimens, respectively, depending on the type of analysis. Similarly, a sub-sample of 30 specimens with a comparable range of $L_{\mathrm{S}}$ (specimens with $103-148 \mathrm{~mm} L_{S}$ ) was chosen from each population.

To test the assumption that the morphological differences result from different temperatures, latitude was used as a surrogate of temperature regimes, as proposed for one of the most successful of the introduced North American nest-guarding species, pumpkinseed Lepomis gibbosus (L. 1758) (Copp et al., 2002; Copp \& Fox, 2007). Thus, to examine the potential latitudinal differences among the four populations, morphometric characters (expressed as the mean values of $\% L_{S}$ ) were sorted following the co-ordinates of each site from north to south. Pearson correlation coefficients $(r)$ were computed and linear regression analysis (least-square method, $P<0.05$ ) was applied.

## RESULTS

## BASIC MORPHOMETRY WITHIN ONTOGENETIC CONTEXT

Specimens of A. melas collected from the U.K. ranged from 60.3 to 193.8 mm $L_{\mathrm{S}}($ mean $=94.5 \mathrm{~mm}$ ), specimens from the French population ranged from 49.1 to $200.8 \mathrm{~mm} L_{\mathrm{S}}($ mean $=130.5 \mathrm{~mm}$ ), Italian $A$. melas ranged from 29.5 to 229.4 mm $L_{\mathrm{S}}($ mean $=158.3 \mathrm{~mm})$ and $A$. melas from Slovakia ranged from 17.6 to 184.9 mm $L_{\mathrm{S}}($ mean $=101 \cdot 8 \mathrm{~mm}$ (Table I).
In the TRAs, A. melas from the U.K. demonstrated isometric development in 17 morphometric characters, allometric development in seven characters and isometric development with an abrupt change in six characters (Table II) that occurred between 177.2 and $190.7 \mathrm{~mm} L_{\mathrm{S}}$ [Fig. 3(a)], i.e. very late in ontogeny. In A. melas from France, the majority (22) of morphometric characters developed isometrically [Fig. 3(b) and Table II], whereas seven characters demonstrated allometric development and only one morphometric character developed isometrically with abrupt change (breakpoint occurring at $158 \mathrm{~mm} L_{\mathrm{S}}$ ). In the Italian A. melas, 12 morphometric characters developed isometrically, 15 allometrically and three isometrically with abrupt changes that occurred between 166.9 and $226.4 \mathrm{~mm} L_{\mathrm{S}}$ [Fig. 3(c) and Table III], i.e. only in adult specimens. In A. melas from Slovakia, 10 characters developed isometrically, seven allometrically and 13 isometrically with abrupt changes that occurred between 77.7 and $159.9 \mathrm{~mm} L_{\mathrm{S}}$ [Fig. 3(d) and Table III).

According to the geometric-shape analysis (allometric form-space PCA), adults in all four populations appeared to have a significantly different body shape than juveniles (Fig. 4). During ontogeny, the head became relatively shorter, the eyes migrated anteriorly, the abdominal area became relatively larger and the caudal fin deepened. The development of body proportions in all four populations, however,


FIG. 3. Estimates of breakpoints and their s.E. (see Tables II and III) in four non-native European populations of Ameiurus melas. Breakpoints in the morphometric characters (listed in Table I) of four non-native European Ameiurus melas populations demonstrate significant abrupt changes in slope when plotted against standard length ( $L_{S}$ ) (Kováč et al., 1999): (a) U.K., (b) France), (c) Italy and (d) Slovakia.
appeared equal and parallel (Fig. 4). Differences between full-model (each of the four populations taken into account separately) and any particular sub-model (all four populations pooled together) revealed that all populations had the same initial shape position $\left(F_{2,805}=1.6933, P>0.05\right)$ and identical developmental slopes, i.e. the same developmental directions $\left(F_{2,805}=3.3862, P>0.05\right)$. In all four populations examined, the influence of PC1 highly predominates ( $96.67 \%$ ). The effect of developmental changes associated with growth dominated, whereas the interpopulation non-allometric variability was almost negligible (only 3.33\%; Fig. 4). Even if the direction of the developmental changes was identical in all populations (changes related to the longitudinal axis of body), their intensity in the developmental trajectories was not. As a result, developmental changes associated with growth led to different overall phenotypes among populations.

No significant difference ( $C_{\text {dif }}$ ) was found in body proportions between subsamples of the 30 smallest specimens from each population. In sub-samples of the 30 largest specimens, however, some statistically significant differences were observed among populations: preorbital distance was smaller in French than in Italian specimens; C-fin length and eye diameter were larger in U.K. than in Italian specimens; eye diameter and C-fin length were significantly larger and preorbital distance significantly shorter in Slovak than in Italian specimens (all differences also visible in Figs 5 and 6). To sum up, juvenile $A$. melas were found to have a relatively uniform body shape regardless of the population's origin, whereas adults developed different phenotypes depending upon location.

## INTER-POPULATION COMPARISONS

The highest similarity in the development of individual distance-based morphometric characters was found between U.K. and French populations, with 17 (predominantly isometric) characters showing the same developmental pattern


Fig. 4. (b) Growth changes during ontogeny in the four populations of Ameiurus melas ( - - U. U.; - - France; $\cdot \cdot+\cdot$; Italy, $\cdot->-$, Slovakia), illustrated on thin-plate spline (TPS) grids of form-space principal component analysis (PCA)-relative warp analysis (RWA) ( $96.67 \%$ ). Form-space PCA (RWA) explained variability is $97.53 \%$. The body shape of (a) immature and (c) adult specimens (both in direction of RW1).
(Fig. 7 and Table II). A different pattern was found in the Italian population, with most characters developing allometrically (Fig. 7 and Table III); whereas in the Slovak population, the majority of characters developed isometrically with abrupt changes [Figs 3(d) and 7 and Table III]. In all examined populations, the breakpoints appeared late in ontogeny, i.e. in adults (Fig. 3 and Tables II and III).

Size-adjusted shape-space PCA of the geometric analysis (Fig. 5), size-adjusted shape space LDA (Fig. 6) and the $C_{\text {dif }}$ results (see above) indicated that Italian $A$. melas, relative to the U.K. and Slovak populations had proportionally smaller and more posteriorly situated eyes, a larger head, anteriorly placed dorsal fin, posteriorly placed adipose fin, shallower caudal peduncle and shorter and smaller caudal fin. The French A. melas represented an intermediate form between the U.K. and Slovak


Fig. 5. (b) Inter-population differences among the four populations of Ameiurus melas ( - , U.K.; - - -, France; $\cdot+\cdots$; Italy, $--* \cdot$, Slovakia), illustrated on thin-plate spline (TPS) grids of size-adjusted shapespace principal component analysis (PCA)-relative warps analysis (RWA). The shape-space PCA (RWA) size-adjusted shape space explained variability is $35.27 \%$. The most discriminative body shape is shown in the (a) Italian and (c) U.K. and Slovak populations of A. melas (both in the direction of RW1).


FIG. 6. (b) Inter-population differences among the four populations of Ameiurus melas (---, U.K.; - - - France; ${ }^{+}+$; Italy, $-x$ - Slovakia), illustrated on thin-plate spline (TPS) grids of size-adjusted shape-space linear discriminant analysis (LDA). Linear discriminant analysis size-adjusted shape space explained variability $=90.44 \%$. The most discriminative body shape is shown in the (a) U.K. and Slovak and (c) Italian populations of A. melas (both in the direction of LD1).
and Italian phenotypes (Figs 5 and 6). In the test of latitudinal effect, a statistically significant difference $(P<0.05)$ was found in two morphometric characters: caudalfin depth and caudal-fin length.

## DISCUSSION

One of the typical attributes of successful invasive fish species appears to be their high morphological variability and phenotypic plasticity (Tomeček et al., 2005; Záhorská et al., 2009), as body shape is directly or indirectly influenced by the ambient environmental conditions (Norton et al., 1995). This has been observed in


Fig. 7. Correspondence analysis of the four populations of Ameiurus melas $(\bullet$ U.K.; $\mathbf{\Delta}$, France; + , Italy; $\times$, Slovakia). Graph shows the predominant type of development of particular morphometric characters in all populations examined in this study and their reciprocal relativeness.
several widespread species of fishes, such as sticklebacks Gasterosteus spp. (Kováč et al., 2002), L. gibbosus (Tomeček et al., 2005) or topmouth gudgeon Pseudorasbora parva (Temminck \& Schlegel 1846) (Záhorská et al., 2009).

Freshwater invasive fishes in newly occupied areas usually tend to attain smaller body size compared to individuals from native populations (Tomeček et al., 2005; Záhorská et al., 2009). This is often caused by modification of their life histories, especially by a shift in allocation of their resources to reproduction and maintenance of offspring rather than to somatic growth (Novomeská et al., 2010). Indeed, individuals from all four European populations of A. melas examined in this study are considerably smaller relative to those from native North American populations, where $A$. melas grows up to $467 \mathrm{~mm} L_{\mathrm{T}}$ or even $610 \mathrm{~mm} L_{\mathrm{T}}$ (Carlander, 1969; Scott \& Crossman, 1973). Individuals from populations with the longest period of introduction, i.e. from the Italian population established in the Lake Trasimeno for at least 40 years (Gianotti et al., 1975) and from the French population established in Brière Marsh for at least 70 years (Cucherousset et al., 2006), show larger body size (both maximum and mean values) compared to more recently established U.K. and Slovak populations (see Table I). Therefore, it might seem that a longer time of establishment gradually leads back to the body size values typical for native populations. Nevertheless, body size in all four populations examined still remains smaller compared to native A. melas. The same phenomenon has been recorded in North America for non-native round goby Neogobius melanostomus (Pallas 1814), which attains a smaller body size than specimens from native Ponto-Caspian populations (MacInnis \& Corkum, 2000), or in L. gibbosus from some non-native European sites that were found to be smaller than L. gibbosus from native North American populations even after 100 years of establishment (Copp et al., 2004; Copp \& Fox, 2007). In this case, the differences are likely to be related to differences in temperature regime, which influences both the amount of food available and timing of somatic growth.

In all populations examined, breakpoints occur only in adult specimens and in most cases quite late in ontogeny. This suggests that A. melas attains its definitive phenotype early in ontogeny and no substantial body shape changes appear later. It may be associated with a relatively uniform life history of $A$. melas throughout its ontogeny from early juvenile to adult periods. In many species, ontogenetic changes in body shape go hand in hand with adaptation to different habitat use (e.g. lentic $v$. lotic and pelagic $v$. benthic) (Tomeček et al., 2005), diet type (insects $v$. larger prey) (Watson \& Balon, 1984; Kováč et al., 1999) or other ecological requirements. In such cases, the abrupt changes in morphometric characters can be considered a potential tool to identify thresholds in the fish's life history (Copp \& Kováč, 1996; Kováč et al., 1999). On the other hand, breakpoints in morphometry can be considered to be developmental thresholds only if they coincide with each other and with other morphological, physiological, ecological or behavioural changes during ontogeny (Kováč et al., 1999). This is not the case in A. melas, however, and the breakpoints found late in its ontogeny do not seem to have any substantial functional meaning, especially if the importance of changes in external morphology decreases with size and age of fish (Kováč et al., 1999).

A previous study of the Slovak population concluded that ontogenetic changes in external morphology of this non-native population occur continuously, and that both juveniles and adults appeared to show little morphological variation between individuals (Novomeská et al., 2010). Such uniformity within the population may
reflect a possible founder effect and low phenotypic plasticity in this population. Similarly, no statistically significant differences (see $C_{\text {dif }}$ results) were found among juveniles when all four non-native populations were compared. Continuous developmental changes in these populations, however, result in increased variation between adult individuals of the same population, as well as between populations. In other words, low morphological variability appears to be restricted to the Slovak population, whereas a pattern of continuous ontogenetic change in external morphology (i.e. without apparent thresholds) appears to be typical for all non-native A. melas populations. Such a pattern was also observed in other introduced species in Europe, e.g. P. parva (Záhorská et al., 2009) or L. gibbosus (Tomeček et al., 2005; Copp \& Fox, 2007), where different adult morphotypes are formed, whereas the morphology of juveniles appear rather uniform across the geographical range of the species (i.e. both in its native and non-native populations). In P. parva, for example, the phenotypic plasticity appears to be much higher than that in A. melas, as it is expressed not only in the formation of different definite phenotypes but also in the manner by which the phenotypes are achieved (Záhorská et al., 2009). Indeed, in A. melas the ontogenetic trajectories were found to be parallel, and the overall morphological variability is affected by interindividual noise rather than by differences between populations. Moreover, high variability in P. parva can result from the very short lifespan of this species, which allows very fast alteration of generations and thus a potential to generate novel phenotypes faster.

The Italian phenotype was found to be the most distinct among the four populations of A. melas (Figs 5-7). In general, variations in external morphology can result from different genetic attributes of the populations, from different environmental conditions, or from the combination of both. Populations of A. melas were imported to Europe (mainly to Italy, Germany, the Netherlands and Belgium) for experimental purposes, aquaculture and stocking ponds in the late 19th and early 20th centuries (Wheeler, 1978; Copp et al., 2005). The fish probably originate from various sites in North America. Different sources of introduction could thus introduce genetic diversity in these non-native European populations of A. melas, with potential effects on external morphology. Unfortunately, no relevant information on genetic differences of $A$. melas in Europe have been available until now; only a preliminary genetic analysis of specimens from Belgium, France and the Netherlands has been undertaken (Verreycken et al., 2010). On the other hand, in L. gibbosus it has been found that only $14 \%$ of morphological variability is caused genetically, while as much as $53 \%$ is induced by environmental conditions (Robinson et al., 2000). Indeed, body shape in fishes can be influenced by various factors such as temperature (Martin, 1949; Beacham, 1990; Šumer et al., 2005), food ratio (Currens et al., 1989) and type of food or feeding mode (Day et al., 1994; Robinson \& Wilson, 1996). Moreover, phenotypic plasticity is regarded to be more beneficial in variable environments than in constant environments, where a single optimal phenotype may be favoured instead of maintaining plasticity in traits (Schlichting \& Pigliucci, 1998). The thermal conditions associated with lower latitudes could be responsible for the distinct patterns in the morphology of the Italian population, as the water temperature in Lake Trasimeno often exceeds $30^{\circ} \mathrm{C}$ in summer (Lorenzoni et al., 2010) and thus differs considerably from the other sites where $A$. melas for this study were sampled.

It has been demonstrated that $A$. melas from non-native European populations have some potential to alter their body shape both within and between populations.

The phenotypic plasticity of A. melas, however, was not found to be as significant as in other invasive fish species, e.g. P. parva or L. gibbosus. The findings of this study suggest that the morphological variability itself is not necessarily essential for invasive success. The invasiveness of $A$. melas is therefore probably favoured by variations in its life-history traits and reproduction variables, together with some behavioural traits, e.g. voracious feeding, parental care (Scott \& Crossman, 1973; Ribeiro et al., 2008) rather than by phenotypic plasticity expressed in external morphology.

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[^1]:    L, linear; Q , quadratic; S, split-linear development; Fit, best fitting model; BP, breakpoint. D , dorsal fin; P , pectoral fin; V , pelvic fin; A , anal fin; C , caudal fin; adip, adipose fin; NS, not significant ( $P>0.05$ ).

